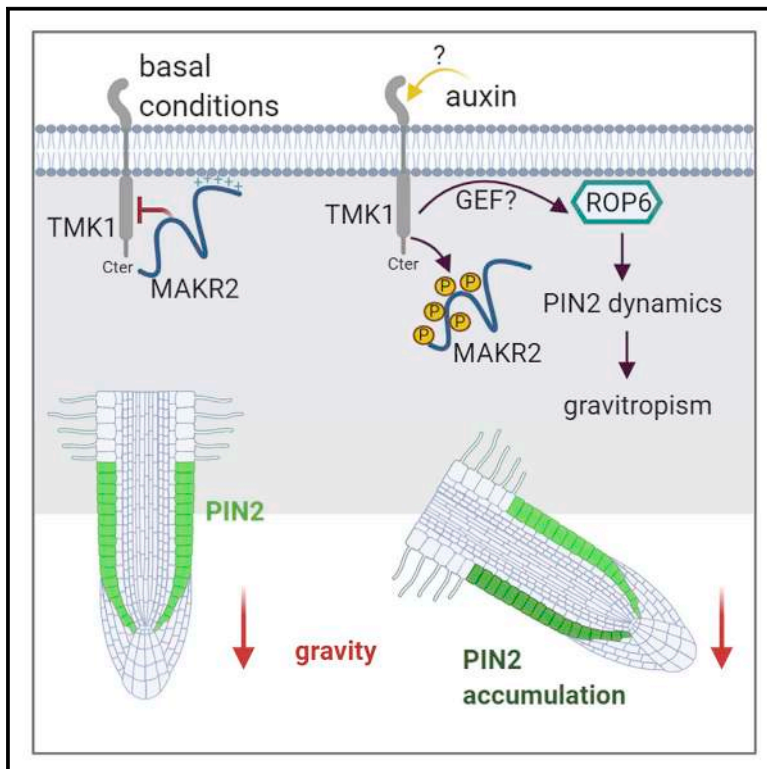


Current Biology

Auxin-Regulated Reversible Inhibition of TMK1 Signaling by MAKR2 Modulates the Dynamics of Root Gravitropism

Graphical Abstract



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In Brief

Marquès-Bueno, Armengot et al. show that the unstructured protein MAKR2 controls the dynamics of the root gravitropic response by acting as a negative regulator of the TMK1 receptor kinase. In addition, the MAKR2 inhibitory effect on TMK1 signaling is antagonized by auxin itself, which triggers rapid MAKR2 membrane dissociation in a TMK1-dependent manner.

Highlights

- MAKR2 is co-expressed with PIN2 and regulates the pace of root gravitropism
- MAKR2 controls PIN2 asymmetric accumulation at the root level during gravitropism
- MAKR2 binds to and is a negative regulator of the TMK1 receptor kinase
- Auxin antagonizes the MAKR2 inhibition of TMK1 by delocalizing MAKR2 in the cytosol



Report

Auxin-Regulated Reversible Inhibition of TMK1 Signaling by MAKR2 Modulates the Dynamics of Root Gravitropism

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<https://doi.org/10.1016/j.cub.2020.10.011>

SUMMARY

Plants are able to orient their growth according to gravity, which ultimately controls both shoot and root architecture.¹ Gravitropism is a dynamic process whereby gravistimulation induces the asymmetric distribution of the plant hormone auxin, leading to asymmetric growth, organ bending, and subsequent reset of auxin distribution back to the original pre-gravistimulation situation.^{1–3} Differential auxin accumulation during the gravitropic response depends on the activity of polarly localized PIN-FORMED (PIN) auxin-efflux carriers.^{1–4} In particular, the timing of this dynamic response is regulated by PIN2,^{5,6} but the underlying molecular mechanisms are poorly understood. Here, we show that MEMBRANE ASSOCIATED KINASE REGULATOR2 (MAKR2) controls the pace of the root gravitropic response. We found that MAKR2 is required for the PIN2 asymmetry during gravitropism by acting as a negative regulator of the cell-surface signaling mediated by the receptor-like kinase TRANSMEMBRANE KINASE1 (TMK1).^{2,7–10} Furthermore, we show that the MAKR2 inhibitory effect on TMK1 signaling is antagonized by auxin itself, which triggers rapid MAKR2 membrane dissociation in a TMK1-dependent manner. Our findings suggest that the timing of the root gravitropic response is orchestrated by the reversible inhibition of the TMK1 signaling pathway at the cell surface.

RESULTS AND DISCUSSION

Receptor-like kinases (RLKs) are involved in all aspects of plant life, including development, immunity, reproduction, and environmental interactions.¹¹ Yet we are still lacking mechanistic details on how those receptors are activated and regulated. Notably, the functions and mechanisms of RLK negative regulation have rarely been addressed, although it is established that receptor inhibition plays a critical role in signaling and diseases in metazoans.¹² BRI1 KINASE INHIBITOR1 (BKI1) is a plant-specific unstructured protein that negatively regulates the activity of the plant steroid receptor BRASSINOSTEROID INSENSITIVE1 (BRI1).^{13–15} BKI1 acts through two evolutionarily conserved linear motifs: a C-terminal alpha helix of 20 residues that binds the BRI1 kinase domain and inhibits the interaction between BRI1 and its co-receptor BRI1 ASSOCIATED KINASE1 (BAK1),^{13–16} and a cationic membrane hook, which targets BKI1 to the plasma membrane.^{14,17} Upon brassinosteroid

perception, BRI1 phosphorylates BKI1 on a conserved tyrosine within the membrane hook,¹⁴ triggering an electrostatic switch that releases BKI1 from the plasma membrane into the cytosol^{17,18} and allowing the transphosphorylation of BRI1/BAK1 kinases and subsequent activation of the pathway.¹⁹

The presence of both the cationic membrane hook and the BRI1-binding peptide at the C terminus defines a novel family of proteins named MEMBRANE ASSOCIATED KINASE REGULATOR (MAKR) composed of 7 members (BKI1 and MAKR1–MAKR6).¹⁴ Like BKI1, these proteins are unstructured cytosolic proteins that are targeted to the plasma membrane via electrostatic interactions.¹⁷ However, with the exception of MAKR1, they are unable to bind to BRI1 kinase and thus they likely control different signaling pathways.^{14,20,21} For example, the auxin-inducible MAKR4 is involved in lateral root formation,²¹ whereas MAKR5 is involved in protophloem differentiation.²⁰ The latter acts as a positive downstream effector of the leucine-rich-repeat (LRR) RLK BARELY ANY MERISTEM3 (BAM3), suggesting that



MAKR proteins may act as positive or negative regulators of RLK signaling.^{20,22}

Here, we addressed the function of MAKR2, a so far uncharacterized member of the MAKR family. We raised *MAKR2* gain- and loss-of-function transgenic *Arabidopsis* lines via overexpression and artificial microRNA (*amiMAKR2*), respectively. To monitor protein accumulation and localization, we fused MAKR2 at its C terminus with either the red fluorescent protein 2xmCHERRY (*2x35S_{prom}::MAKR2-2Ch*, hereafter designated as *MAKR2-Ox1*) or the yellow fluorescent protein mCITRINE (*2x35S_{prom}::MAKR2-mCit*, hereafter designated as *MAKR2-Ox2*). Quantitative RT-PCR and confocal analyses of tagged proteins confirmed the overexpression of *MAKR2* in *MAKR2-Ox* lines and its downregulation in each independent artificial microRNA line (Figures 1A, S1A, and S1C). The roots overexpressing *MAKR2* did not grow vertically, as manifested by an increased gravitropic index (Figure 1B). Analysis of the synthetic auxin output reporter *DR5_{prom}::GUS* showed an arrow-like pattern of GUS accumulation at the root tip of *MAKR2-Ox1* lines with an increased signal on both flanks of the lateral root caps (Figure 1C). This DR5 arrow-like pattern at the root tip is typically observed in *pin2* loss-of-function alleles.^{23,24} Consistent with a potentially impaired PIN-FORMED2 (PIN2) activity, *MAKR2* overexpression inhibited the establishment of the asymmetric patterns observed after 5 h of gravistimulation in both *DR5_{prom}::GUS* and *DR5_{prom}::GFP* reporter lines (Figures 1C and S1B). Quantitative analyses of root bending following gravistimulation showed that *MAKR2-Ox* lines reoriented slowly to the new gravity vector (Figure 1D). In contrast, three independent *amiMAKR2* lines had the opposite phenotype, displaying fast gravitropic bending (Figures 1D and S1C). To validate the specificity of our *amiMAKR2* lines, we created a *crispr* allele (*makr2-1*). This allele led to a truncated protein of 37 residues, comprising only the first 8 residues of MAKR2 and an additional 29 random residues. The *amiMAKR2* lines and the *makr2-1* allele had identical gravitropic phenotypes (Figure S1D). Both the *MAKR2-Ox* and *amiMAKR2.1* lines had slightly shorter primary roots than the wild type (Figure S1E); however, they had opposite gravitropic phenotypes (Figure 1D). Furthermore, we found no correlation between root length (Figure S1E) and the strength of the agravitropic phenotypes of the *MAKR2-Ox1* and *MAKR2-Ox2* lines (Figure 1D). Together, these data suggest that primary root growth is unlikely to explain the gravitropic phenotypes of *MAKR2* gain- and loss-of-function mutants.

The pace of root gravitropism is regulated by Rho GTPase of Plants 6 (ROP6), with the roots of *ROP6* loss-of-function mutants responding slowly to gravistimulation and the roots of *ROP6* gain-of-function mutants (e.g., *ROP6-Ox*) responding faster than wild-type roots.^{8,9,25} Because the loss- and gain-of-function mutants of both *MAKR2* and *ROP6* have opposite phenotypes, we hypothesized that MAKR2 may act as a negative regulator of the ROP6 signaling pathway. To test this, we first assessed whether *MAKR2* is expressed in the same tissues as *ROP6* using a transcriptional reporter line, *MAKR2_{prom}::VENUS^{NLS}*, and translational fusion lines, *MAKR2_{prom}::MAKR2-tdYFP*, *MAKR2_{prom}::MAKR2-GUS*, and *ROP6_{prom}::mCitrine-ROP6*. *MAKR2* was expressed in the root meristem in the epidermis and cortex cell layers, as well as in the quiescent center and surrounding initials (Figures 2A and S2A–S2C). As

reported with the GFP-ROP6 reporter lines,^{8,26,27} we found that our mCit-ROP6 reporter was expressed in the root tip epidermis and cortex cells, albeit to a lower level than in internal tissues (Figure 2B). Therefore, *MAKR2* and *ROP6* expression partially overlap. Next, we addressed the genetic relationship between *ROP6* and *MAKR2* by crossing a *ROP6-Ox* line with our *MAKR2-Ox1* line, which show fast- and slow-gravitropic responses, respectively. *ROP6-Ox;MAKR2-Ox1* double transgenics showed a wild-type-like response, suggesting that *ROP6* overexpression mitigates the strong agravitropic phenotype induced by *MAKR2* overexpression (Figure 2C).

ROP6 has been proposed to mediate root gravitropic bending by regulating the trafficking of the auxin-efflux carrier PIN2.^{2,4,8,9} In particular, ROP6 mediates PIN2 accumulation in the epidermis on the gravity-stimulated side of the root (lower side of the root, facing the new gravity vector).² We therefore analyzed whether *MAKR2* could control PIN2 dynamics during gravitropism. First, we raised a *MAKR2_{prom}::MAKR2-2Ch* line and crossed it with a *PIN2_{prom}::PIN2-GFP* line to monitor their co-expression and localization. We found that *MAKR2* and *PIN2* were co-expressed and co-localized at the plasma membrane of root epidermis and cortex cells (Figure S2D). Next, we crossed the *MAKR2-Ox1* and *amiMAKR2.1* lines with the *PIN2_{prom}::PIN2-GFP* reporter and followed PIN2-GFP accumulation during gravitropism using a vertical stereomicroscope setup.²⁸ High-resolution time-lapse analyses validated the slow- and fast-gravitropic response of *MAKR2-Ox1;PIN2_{prom}::PIN2-GFP* and *amiMAKR2;PIN2_{prom}::PIN2-GFP*, respectively (Figure S2E; Videos S1 and S2). Using time-lapse analysis, we noticed that each root had slightly different gravitropic dynamics (Figure S2E), which made it difficult to pool the quantification of PIN2-GFP fluorescence between replicates. We thus decided to trace PIN2-GFP fluorescence in individual roots shown as independent replicates in Figure 2D. Importantly, although we observed some root-to-root variations in PIN2-GFP dynamics, the overall trend was nevertheless robust from replicate to replicate (Figure 2D). Quantitative measurements of PIN2-GFP fluorescence on the upper and lower sides of the root confirmed that PIN2-GFP is retained on the lower side of the root longer than on the upper side (Figure 2D; Videos S1 and S2).^{2,29,30} In particular, PIN2-GFP signal at the lower part of the wild-type root initially increased before decreasing (Figure 2D). Such signal increase was not observed at the upper part of the root, which instead showed a steady decrease of fluorescence (Figure 2D). The differential PIN2-GFP accumulation between the upper and lower parts of the root was abolished in the *MAKR2-Ox1*-overexpressing line (Figure 2D; Video S1). By contrast, PIN2-GFP accumulation at the lower part of the root in the *amiMAKR2* line was more pronounced and lasted longer than in the wild type (Figure 2D; Video S2). Together, these results indicate that *MAKR2*, like *ROP6*, is required for dynamic PIN2 accumulation during root gravitropism, which could explain the relative gravitropic phenotypes of the *MAKR2-Ox* and *amiMAKR2* lines.

We next investigated whether *MAKR2* may also regulate the activity of an RLK upstream of ROP6 activation. In the context of pavement-cell-shape morphogenesis, ROP6 acts downstream of the LRR RLKs from the TRANSMEMBRANE KINASE (TMK) family, which were proposed to operate as a relay for perception of extracellular auxin.^{2,7,31} We thus wondered

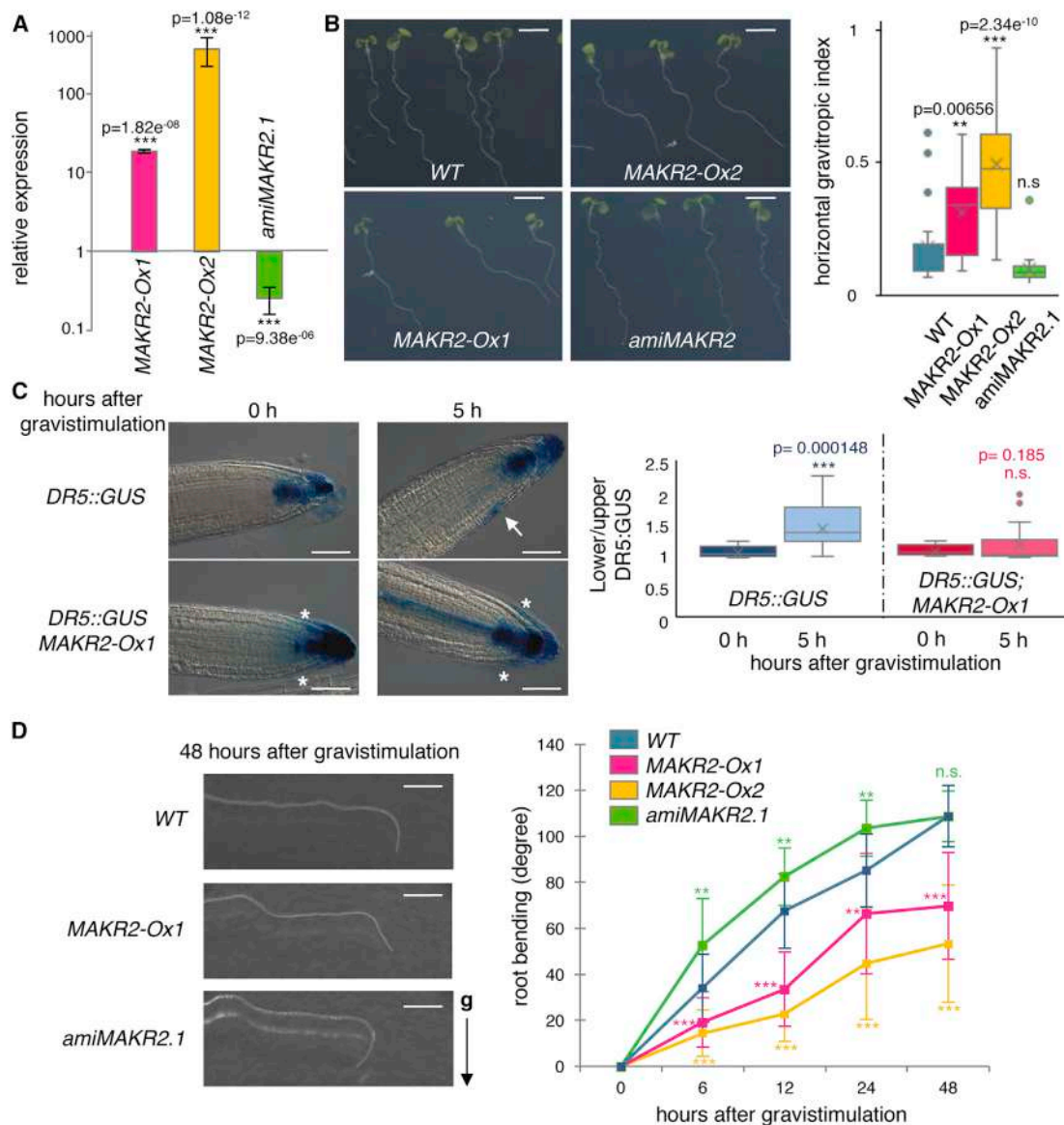


Figure 1. MAKR2 Regulates the Pace of the Root Gravitropic Response

(A) qRT-PCR analyses of *MAKR2* expression in *2x35S_{prom}::MAKR2-2xmCherry* (*MAKR2-Ox1*), *2x35S_{prom}::MAKR2-mCitrine* (*MAKR2-Ox2*), and *amiMAKR2.1* lines relative to *MAKR2* expression in wild-type seedlings (mean \pm SEM).

(B) Pictures showing the root phenotypes of the genotypes indicated at the bottom and related quantification of the horizontal growth index (Tukey boxplot). Plants were grown at a 45° angle with respect to the vertical axis. Scale bars represent 5 mm.

(C) *DR5_{prom}::GUS* accumulation pattern in the absence and after 5 h of gravistimulation at a 135° angle in wild-type and *MAKR2-Ox1* plants and related quantification (Tukey boxplot). The white asterisks indicate the arrow-like pattern observed in *MAKR2-Ox1* lines; the white arrow indicates the asymmetric GUS signal observed after gravistimulation in the wild type. Scale bars represent 50 μ m.

(D) Representative pictures of the root gravitropic curvature 48 h after reorienting seedlings at a 135° angle and related quantification of root gravitropic bending over time (mean \pm SEM). Scale bars represent 2 mm. See Figure S1F for a statistical comparison.

For the horizontal gravitropic index and the kinetics of the gravitropic response, a linear model was fitted on measurements from wild-type plants and the different mutants using *lm()* function from stats package available in R software (<https://www.r-project.org/>). This model estimates a weight for each variable (wild-type and mutant plants) and the associated probability that such weight is different from zero based on a t test. The probability derived from the t test is the p value in this comparison and significant differences were considered when $p < 0.05$. See also Figure S1.

whether TMKs may also participate in root gravitropism. TMKs form a family of redundant receptors, with single mutants having no or subtle root phenotypes,³² whereas higher-order mutants have strong pleiotropic developmental defects.³³ The *tmk1;tmk4*

double mutant showed a reduced root gravitropic response (Figure S3A). Although consistent with the notion that TMK receptors may be involved in root gravitropic bending, the *tmk1;tmk4* mutant also had stunted root growth,³³ making it difficult

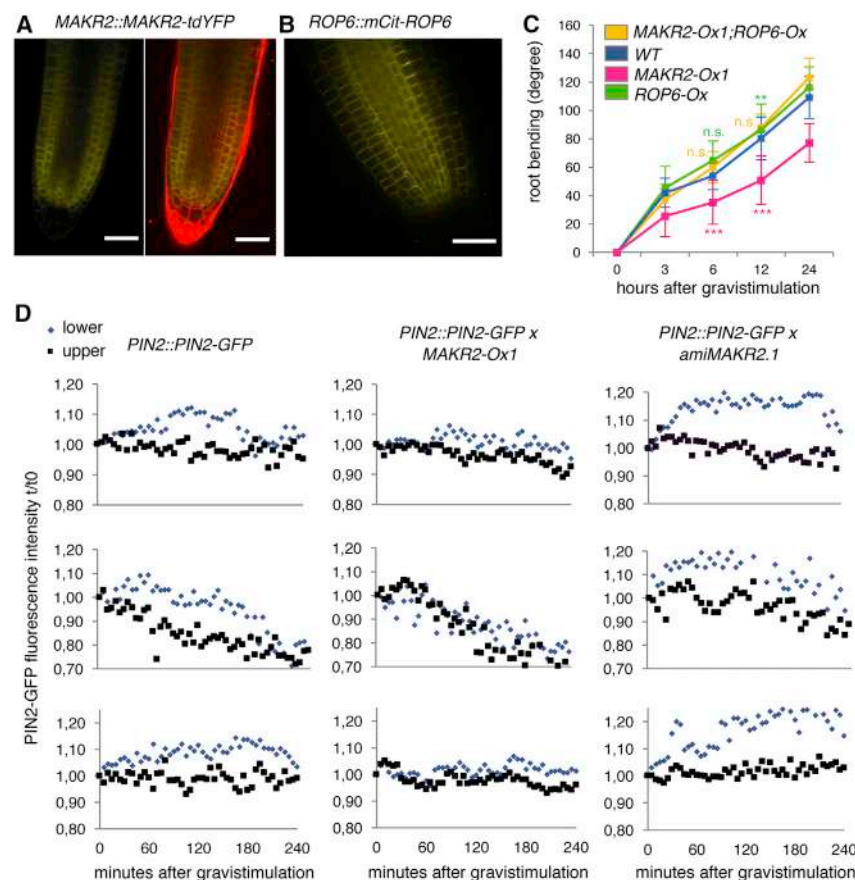


Figure 2. MAKR2 Mediates PIN2-GFP Dynamic Accumulation during Gravitropism

(A) Confocal pictures of the *MAKR2prom::MAKR2-tdYFP* line showing the MAKR2-tdYFP localization and expression pattern at the root tip. Left: yellow fluorescent protein (YFP) channel; right: overlay between YFP channel (yellow) and membranes counterstained by FM4-64 (red).

(B) Confocal pictures of the complemented *ROP6prom::mCitrine-ROP6/rop6-2* line showing the mCit-ROP6 localization and expression pattern at the root tip.

(C) Kinetics of root gravitropic bending after reorienting seedlings at a 135° angle. See Figure S1F for a statistical comparison. A linear model was fitted on measurements from wild-type plants and the different mutants using *lm()* function from stats package available in R software (<https://www.r-project.org/>). This model estimates a weight for each variable (wild-type and mutant plants) and the associated probability that such weight is different from zero based on a t test. The probability derived from the t test is the p value in this comparison and significant differences were considered when $p < 0.05$.

(D) Quantification of PIN2-GFP in the upper (blue diamonds) and lower (black squares) part of the root in the *PIN2prom::PIN2-GFP*, *PIN2prom::PIN2-GFP;MAKR2-Ox1*, and *PIN2prom::PIN2-GFP;amiMAKR2.1* lines. Each graph shows the response in a single individual root (see also Videos S1 and S2). In each case, fluorescence intensities were normalized with respect to the initial fluorescence value (time 0 min).

Scale bars represent 30 μ m.

See also Figures S1 and S2 and Videos S1 and S2.

to conclude whether its gravitropic phenotype was a primary or secondary phenotype. A transcriptional reporter line, *TMK1prom::2Ch^{NLS}*, confirmed that *TMK1* is expressed in all the tissues of the root meristem, including the epidermis and cortex, where *MAKR2*, *PIN2*, and *ROP6* are also expressed (Figure S3B). We therefore analyzed the phenotype of *TMK1* overexpression by generating a *TMK1-2xmCherry* line driven under the control of the ubiquitous promoter of the *UBQUITIN10* gene (*UBQ10prom::TMK1-2Ch*, hereafter referred to as *TMK1-Ox*). Similar to the *ROP6-Ox* and *amiMAKR2* lines, the *TMK1-Ox* line displayed a fast root gravitropic response (Figure 3A). Together, the *tmk1;tmk4* loss-of-function and *TMK1-Ox* gain-of-function phenotypes suggest that *TMK1* may act upstream of *ROP6* signaling both for pavement cell morphogenesis in leaves and in root gravitropism. Interestingly, *TMK1-Ox*; *MAKR2-Ox2* double transgenics had a wild-type-like root gravitropic response (Figure 3A). This genetic analysis shows that overexpression of *MAKR2* is able to counteract the phenotypic effects of *TMK1* overexpression, suggesting that *MAKR2* may act as a negative regulator of *TMK1*, upstream of *ROP6* activation.

We next tested whether *MAKR2* interacts with *TMK1*. We found that Flag-Halo-*MAKR2* directly interacted with the isolated cytosolic domain of *TMK1* *in vitro* (*TMK1^{cyt}*-HA; hemagglutinin), whereas Flag-Halo-mCitrine did not (Figure 3B). This interaction was specific for the cytosolic domain of *TMK1*, as the cytosolic

domain of HAESA-LIKE1 (*HSL1^{cyt}*-HA), which has a related kinase domain to *TMK1*, did not interact with Flag-Halo-*MAKR2* (Figure S3D). Furthermore, full-length *TMK1* (*TMK1-3HA*), but not a kinase-deleted version (*TMK1 Δ kinase-3HA*), co-immunoprecipitated with *MAKR2*-mCit when co-expressed in protoplasts (Figure 3C). One hour of auxin treatment (1 μ M indole 3-acetic acid; IAA) did not have a strong effect on the interaction between *TMK1-3HA* and *MAKR2*-mCit in this protoplast assay (Figure 3C). Next, we tested whether *MAKR2* interaction with *TMK1* was dependent upon its kinase activity and whether *TMK1* may phosphorylate *MAKR2*. To this end, we co-expressed in bacteria HA-*MAKR2* with the isolated cytosolic domain of *TMK1* in its wild-type or kinase-dead form (6His-Flag-*TMK1^{cyt}* and 6His-Flag-*TMK1^{cyt}*-K616R, respectively). We purified both *TMK1* proteins on a nickel column and identified potential interacting proteins by mass spectrometry. We found *MAKR2* to be the only protein represented by more than one peptide to co-purify with both active and inactive *TMK1* kinase domains. In each case, we recovered 13 unique peptides in *MAKR2*, representing a 42% peptide coverage (63% coverage for both *TMK1^{cyt}* and *TMK1^{cyt}*-K616R) (Figure 3D). These results indicate that *MAKR2* directly interacts with the kinase domain of *TMK1* *in vitro*, irrespective of its kinase activity. We next assessed the phosphorylation status of each peptide recovered during the mass spectrometry experiments. Such analysis of the phosphosites revealed that the *MAKR2* C terminus was phosphorylated

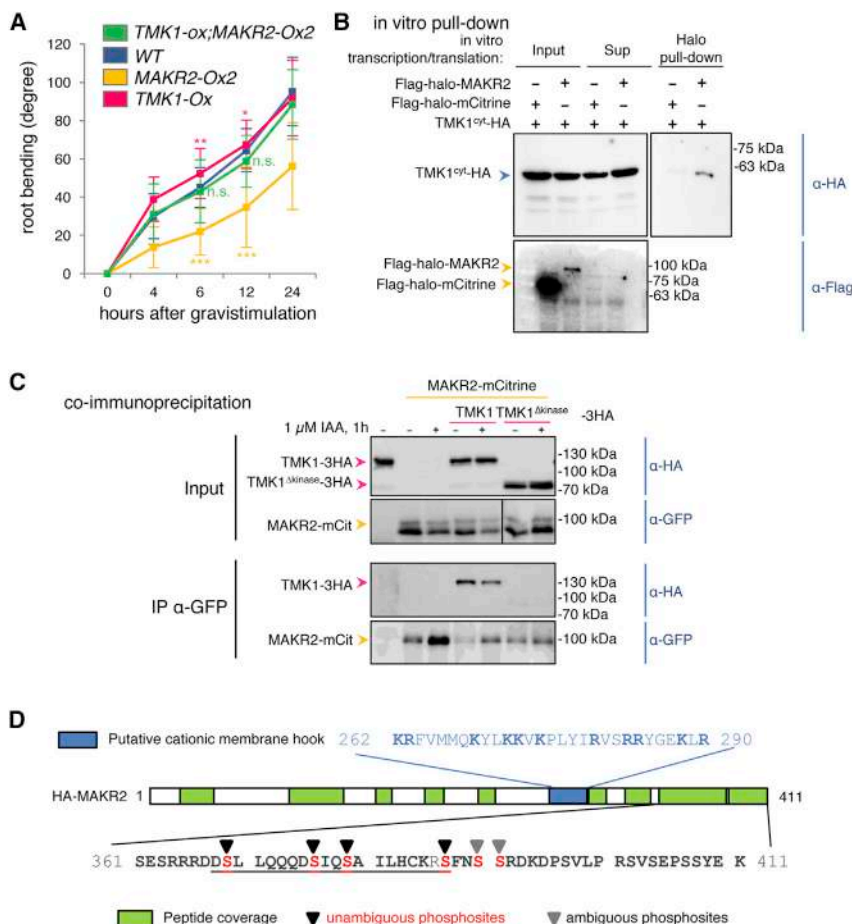


Figure 3. TMK1 Interacts with and Phosphorylates MAK2 and Acts Upstream of MAK2 in the Regulation of Root Gravitropism

(A) Kinetics of root gravitropic bending after reorienting seedlings of the genotypes indicated in the top left corner at a 135° angle. See Figure S3C for a statistical comparison. A linear model was fitted on measurements from wild-type plants and the different mutants using `lm()` function from stats package available in R software (<https://www.r-project.org/>). This model estimates a weight for each variable (wild-type and mutant plants) and the associated probability that such weight is different from zero based on a t test. The probability derived from the t test is the p value in this comparison and significant differences were considered when $p < 0.05$.

(B) Pull-down assay using *in-vitro*-transcribed/translated proteins and Halo-tag purification. Copurified proteins were visualized using an anti-HA antibody (labeled Halo pull-down). The inputs (labeled Inputs) and supernatant (labeled Sup) were tested to show the relative amounts of Halo- and HA-tagged proteins and the binding efficiency to HaloLink magnetic beads (as described in Yazaki et al.³⁴). TMK1^{cyt} corresponds to the isolated TMK1 cytoplasmic domain.

(C) Co-immunoprecipitation of full-length TMK1-3HA but not TMK1^{kinase}-3HA with MAK2-mCitrine. Immunoprecipitation (IP) of MAK2-mCitrine with an anti-GFP antibody and immunoblotting (IB) using an anti-GFP antibody or anti-HA antibody. Protoplasts were incubated or not for 1 h with 1 μM IAA.

(D) The scheme represents the MAK2 protein, with the peptides recovered by mass spectrometry highlighted in green and the phosphorylation sites

shown by arrowheads (only found with active TMK1^{cyt} but not inactive TMK1^{cyt-K616R}). Black arrowheads indicate phosphorylation sites that could be determined with 100% accuracy, whereas gray arrowheads indicate ambiguity on which of the two consecutive serines is phosphorylated. The residues corresponding to the conserved C-terminal tail are underlined. The blue box indicates the position of the putative cationic membrane hook, and the corresponding Arg/Lys residues are highlighted in bold.

See also Figure S3.

on 5 serines when it was co-expressed and co-purified with TMK1^{cyt} (Figure 3D). By contrast, we found no phosphorylation sites when MAK2 was co-expressed and co-purified with TMK1^{cyt-K616R}. Altogether, our results suggest that MAK2 directly interacts with and is phosphorylated by TMK1 *in vitro*, and interacts with full-length TMK1 *in vivo*.

MAK2prom::MAK2-tdYFP expression was too low to follow its localization during the gravitropic response, which provides an endogenous auxin treatment on the lower (i.e., gravistimulated) side of the root. As an alternative, we followed MAK2 localization after exogenous auxin treatments. We found a time- and dose-dependent effect of the synthetic auxin 1-naphthaleneacetic acid (NAA), which induced the relocalization of MAK2prom::MAK2-tdYFP into the cytosol as early as 5 min post treatment (Figure 4A). This rapid effect of auxin treatment was also observed after 5 min of IAA application (Figure 4A). In addition, microfluidics, coupled with time-lapse imaging of MAK2prom::MAK2-tdYFP localization, confirmed that this effect was rapid (i.e., within 2 min following either NAA or IAA treatment) (Figures 4B and S4A; Video S3).³⁵ MAK2 released into the cytosol was independent of protein translation, as shown

by pretreatment with the protein synthesis inhibitor cycloheximide (CHX) (Figure S4B). Furthermore, inhibition of protein degradation by the proteasome inhibitor MG-132, which is required for auxin-mediated gene expression regulation by the TIR1 family,³⁶ had no effect on auxin-triggered MAK2 plasma membrane dissociation (Figure S4B). This suggested that the MAK2 plasma-membrane-to-cytosol localization switch was regulated by a potential “non-transcriptional” arm of auxin signaling at or close to the plasma membrane, rather than by a TIR1-mediated regulation of transcription.² Accordingly, the auxin antagonist PEO-IAA, known to inhibit TIR1-mediated nuclear activity but not the non-transcriptional arm of auxin signaling,^{29,37} efficiently displaced MAK2 away from the plasma membrane (Figure S4C). By contrast, 5-F-IAA, an auxin analog that is able to activate TIR1-mediated gene expression but not the ROP6 pathway,^{29,37} had little effect on MAK2 localization (Figure S4C). Furthermore, benzoic acid (BA), an inactive auxin analog with a pKa similar to that of NAA, and brassinolide had no effect on MAK2prom::MAK2-tdYFP localization (Figure S4D). To address whether this auxin effect was dependent upon the TMK receptors and to bypass the problem due to

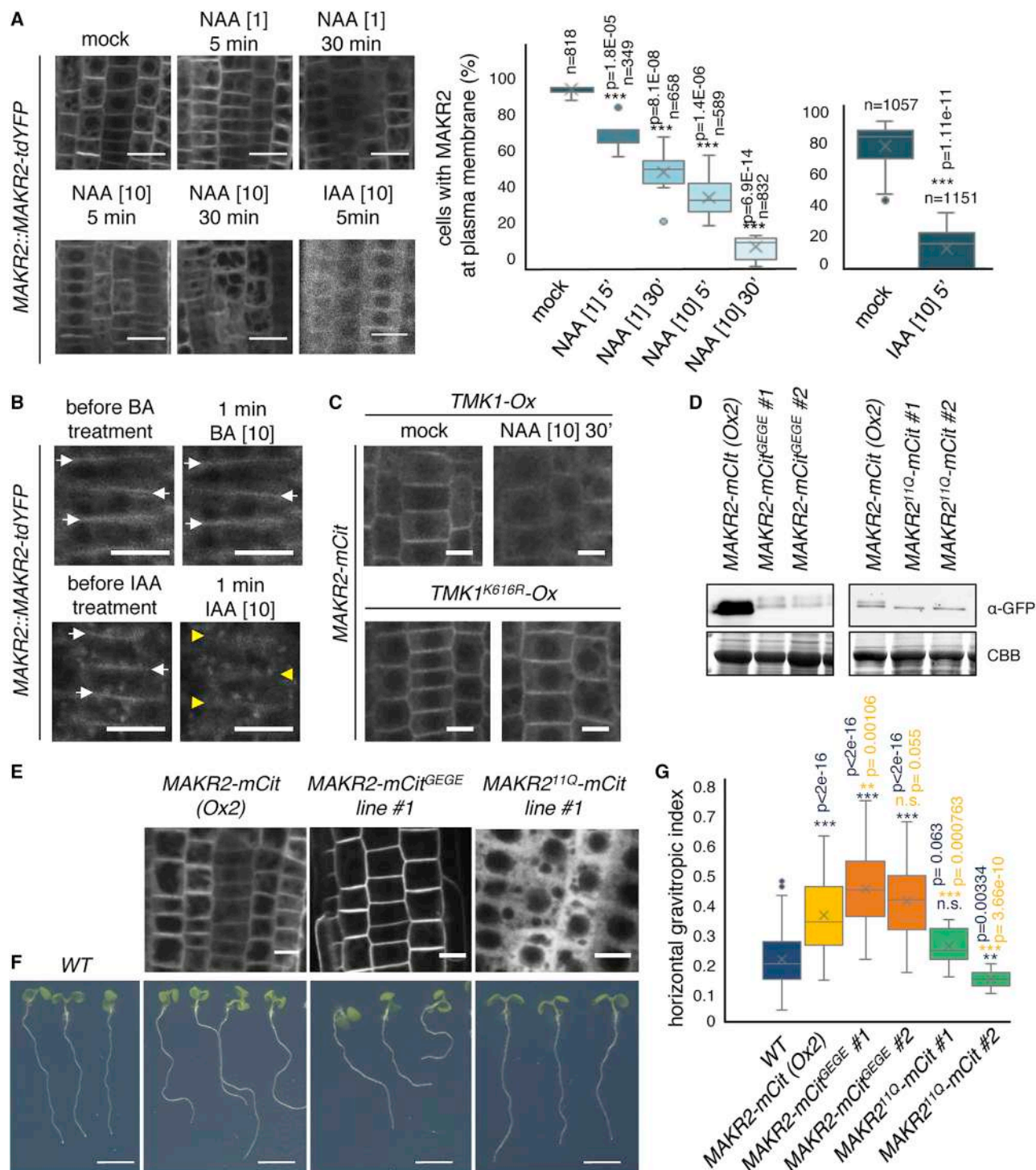


Figure 4. Auxin Triggers MAK2 Plasma Membrane Dissociation in a TMK1-Dependent Manner to Antagonize MAK2 Inhibitory Activity
(A) Confocal pictures of the *MAK2prom::MAK2-tdYFP* line following NAA or IAA treatment for the time and concentration indicated in each panel and related quantification. n indicates the number of cells counted. A pairwise comparison between mock plants and plants subjected to different treatments was performed using a t test with Welch's correction to account for unequal variances using R software (<https://www.r-project.org/>). The probability derived from the t test is the p value in this comparison and significant differences were considered when $p < 0.01$.
(B) Successive confocal pictures of the *MAK2prom::MAK2-tdYFP* line before and after 1 min of benzoic acid (BA; control) or IAA treatment (Video S3). White arrows indicate MAK2 plasma membrane localization, whereas the yellow arrowheads show MAK2 disappearance from the plasma membrane upon IAA but not BA treatment.

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functional redundancy between the four TMK family members, we used a dominant-negative strategy, by overexpressing a kinase-dead version of full-length TMK1 (TMK1^{K616R}).^{7,38} TMK1^{K616R} overexpression severely impaired the auxin-induced MAK2-mCit release from the plasma membrane, whereas overexpression of the wild-type version of TMK1 did not (Figures 4C and S5A). Note that TMK1^{K616R}-mCH-overexpressing plants had a mosaic expression and that the inhibition of MAK2-mCit released from the plasma membrane into the cytosol following auxin treatment was only observed in cells expressing TMK1^{K616R}-mCh (Figure S5B). Together, these results suggest that auxin rapidly regulates MAK2 localization, via a non-transcriptional, TMK-dependent mechanism.

We next addressed the functional impact of the MAK2 localization switch on root gravitropism. To this end, we first quantified the relative plasma membrane and cytosolic localization of MAK2 in the *MAK2-Ox1* and *MAK2-Ox2* lines (Figure S5C). We found that MAK2 in *MAK2-Ox1*, which is less expressed than in the *MAK2-Ox2* line (Figure 1A), had a more pronounced localization in the cytosol and accumulated less at the plasma membrane (Figure S5C). This result is consistent with a model in which MAK2 inhibits the activity of TMK1, which itself triggers the relocalization of MAK2 in the cytosol. Indeed, according to this model, a strong *MAK2* overexpression would lead to a strong inhibition of TMK1 activity, which would not be able to induce the efficient release of MAK2 into the cytosol. By contrast, a mild overexpression of *MAK2* would lead to a milder inhibition of TMK1, which would be able to trigger more MAK2 release from the plasma membrane into the cytosol. To further test this model, we engineered MAK2 mutant versions that are constitutively localized at the plasma membrane or in the cytoplasm. To lock MAK2 at the plasma membrane, we added a C-terminal geranylgeranylation sequence (MAK2-mCit^{GEGE}). MAK2-mCit^{GEGE} was exclusively found at the plasma membrane, by comparison with MAK2-mCit, which localized both to the plasma membrane and in the cytosol (Figure 4E). Lines overexpressing *MAK2-mCit^{GEGE}* had a strong agravitropic phenotype, similar to or even stronger than the *MAK2-Ox2* lines (Figures 4F and 4G), whereas their expression levels were much lower (Figure 4D). This result indicates that a MAK2 protein constitutively localized at the plasma membrane is extremely potent to inhibit root gravitropism. We previously showed that MAK2 interacts with anionic lipids *in vitro* and relies on plasma membrane electrostatics for plasma membrane localization in yeast and *Arabidopsis*.¹⁷ To inhibit MAK2 membrane-binding ability, we mutated the 11 lysine and arginine residues within its

putative membrane hook (Figure 3D) into neutral glutamines (MAK2^{11Q}-mCit). MAK2^{11Q}-mCit was cytosolic (Figure 4E), confirming that MAK2 likely localizes to the plasma membrane through electrostatic interactions with anionic lipids. Importantly, unlike *MAK2-Ox2*, *MAK2^{11Q}-mCit*-overexpressing lines did not have an elevated horizontal gravitropic index even when expressed at similar levels (Figures 4D–4G). Together, these results suggest that MAK2 inhibits gravitropism at the plasma membrane, likely through its interaction with the TMK1 receptor, and that auxin, via the activity of the TMK1 receptor itself, antagonizes this inhibition by triggering the relocalization of MAK2 into the cytosol.

Altogether, our results suggest that MAK2 acts as an upstream negative regulator of ROP6 during the gravitropic response by directly interacting with the TMK1 kinase domain. However, the exact mechanism by which TMK1/MAK2 regulate ROP6 activity remains unresolved. Both TMK1 and ROP6 have been shown to be involved in fast non-transcriptional auxin response,^{7–9,25,32} although the exact mode of auxin perception at or near the cell surface is still unclear.² We propose that the MAK2-mediated negative regulation of TMK1 activity is counterbalanced by auxin itself, which triggers MAK2 relocalization from the plasma membrane into the cytosol. It is possible that this relocalization is induced by TMK1-mediated phosphorylation. However, it is unlikely that the phosphorylation sites that we uncovered in the MAK2 C-terminal tail are directly involved in the regulation of MAK2 localization, because they are far away from the MAK2 membrane hook and their exact function remains to be experimentally determined.

Although BK1 and MAK5 appear to essentially work as inhibitor and activator, respectively, of the BRI1 and BAM3 signaling pathways,^{15,20} our results suggest a parallel mode of action of MAK2 and BK1 in downregulating TMK1 and BRI1 signaling at the cell surface, respectively. We propose that MAK2/TMK antagonist activity allows finely tuning ROP6 signaling during the gravitropic response and thereby regulates the timing of root bending in response to gravity. Together, our results emphasize the importance of RLK negative regulation, which appears critical to determine the strength and dynamics of the output signal.

STAR★METHODS

Detailed methods are provided in the online version of this paper and include the following:

(C) Confocal pictures showing MAK2-mCit localization (*MAK2-Ox2*) in *UBQ10prom::TMK1-2xmCherry* (TMK1-Ox) and *UBQ10prom::TMK1^{K616R}-2xmCherry* (TMK1^{K616R}-Ox, kinase dead) in the absence or presence of NAA at the indicated time and concentration.

(D) Anti-GFP western blots showing the relative accumulation of MAK2-mCit (in the *MAK2-Ox2* line) and two independent transgenic lines overexpressing MAK2-mCit^{GEGE} and MAK2^{11Q}-mCit. CBB, Coomassie brilliant blue.

(E) Confocal pictures comparing the localization of *2x35Spro::MAK2-mCit* (*MAK2-Ox2*), *2x35Spro::MAK2-mCit^{GEGE}* (MAK2-mCit^{GEGE}, constitutively tethered to the plasma membrane), and *2x35Spro::MAK2^{11Q}-mCit* (MAK2^{11Q}-mCit, constitutively cytoplasmic).

(F and G) Pictures showing the root phenotypes of the genotypes indicated at the top (F) and related quantification of the horizontal gravitropic index (G). Statistical comparison with the wild type (WT) is in blue and with *MAK2-Ox2* is in yellow. In (G), a linear model was fitted on measurements from wild-type plants and the different mutants using lm() function from stats package available in R software (<https://www.r-project.org/>). This model estimates a weight for each variable (wild-type and mutant plants) and the associated probability that such weight is different from zero based on a t test. The probability derived from the t test is the p value in this comparison and significant differences were considered when p < 0.05.

Scale bars represent 20 μm (A), 10 μm (B, C, and E), and 5 mm (F).

See also Figures S4 and S5 and Video S3.

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SUPPLEMENTAL INFORMATION

Supplemental Information can be found online at <https://doi.org/10.1016/j.cub.2020.10.011>.

ACKNOWLEDGMENTS

We thank the SiCE group for discussions and comments; S. Yalovsky, B. Scheres, and the NASC/ABRC collection for providing transgenic *Arabidopsis* lines and plasmids; L. Kalmbach and M. Barberon for the gift of *pLOK180_pFR7m34GW*; A. Lacroix, J. Berger, and P. Bolland for plant care; and M. Fendrych for help with microfluidics in the J.F. lab. We acknowledge the contribution of the SFR Biosciences (UMS3444/CNRS, US8/Inserm, ENS de Lyon, UCBL) facilities: C. Lionet, E. Chatre, and J. Brocard at LBI-PLATIM-MICROSCOPY for assistance with imaging, and V. Guegen-Chaignon and A. Page at the Protein Science Facility (PSF) for assistance with protein purification and mass spectrometry. Y.J. was funded by ERC grant 3363360-APPL under FP/2007–2013. Y.J. and Z.L.N. were funded by an ANR- and NSF-supported ERA-CAPS project (SICOPID: ANR-17-CAPS-0003-01/NSF PGRP IOS-1841917). A.I.C.-D. is funded by an ERC consolidator grant (ERC-2015-CoG-683163) and BIO2016-78955 grant from the Spanish Ministry of Economy and Competitiveness. Exchanges between the Y.J. and T.B. laboratories were funded by Tournesol grant 35656NB. B.K.M. was funded by the Omics@vib Marie Curie COFUND and Research Foundation Flanders for a postdoctoral fellowship.

AUTHOR CONTRIBUTIONS

M.M.M.-B., L.A., and M.L. performed cloning and raised and characterized the genetic materials; M.M.M.-B., L.A., and L.C.N. quantified gravitropic bending and performed western blot analyses of protein accumulation in MAKR2 mutant lines; M.M.M.-B. performed all microscopy experiments (double-blind quantifications in collaboration with L.A.); M.M.M.-B. performed DR5::GUS and DR5::GFP analyses in A.I.C.-D.'s laboratory; M.M.M.-B. performed the kinematic analysis of PIN2-GFP during gravitropism in T.B.'s laboratory (with the help of D.O., B.K.M., and S.V.); M.M.M.-B. performed the microfluidics experiments in J.F.'s laboratory; J.B. and L.A. performed *in vitro* Halo-pull down and co-expression/purification in bacteria and related cloning; L.A. designed, isolated, characterized, and phenotyped the *makr2-1* crisper allele with the assistance of Z.L.N.; L.R. performed co-immunoprecipitation in J.F.'s laboratory; M.P.P. provided genetic materials; V.B. assisted with microscopy and image

quantification; Y.J. supervised research; M.M.M.-B., L.A., and Y.J. conceived the study, designed the experiments, and wrote the manuscript. All the authors discussed the results and commented on the manuscript.

DECLARATION OF INTERESTS

The authors declare no competing interests.

Received: June 27, 2019

Revised: September 4, 2020

Accepted: October 7, 2020

Published: November 5, 2020

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STAR★METHODS

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
Mouse monoclonal anti-Flag clone M2	Sigma-Aldrich	Cat#1804; RRID: AB_262044
Mouse monoclonal anti-HA clone 12CA5	Sigma-Aldrich	Cat#11583816001; RRID: AB_514505
Mouse monoclonal anti-GFP HRP clone GG4-2C2.12.10	Miltenyi Biotec	Cat#130-091-833; RRID: AB_247003
Rat monoclonal anti-HA High Affinity clone 3F10	Sigma-Aldrich	Cat#11867423001; RRID: AB_390918
Bacterial and Virus Strains		
<i>Agrobacterium tumefaciens</i> C58 GV3101	N/A	N/A
<i>Escherichia coli</i> DH5 α pa	N/A	N/A
<i>Escherichia coli</i> BL21 (DE3) Rosetta 2 Novagen	Sigma-Aldrich	Cat#69450
Chemicals, Peptides, and Recombinant Proteins		
Cycloheximide (CHX)	Sigma-Aldrich	Cat#C1988
MG-132	VWR	Cat#474790-5
NAA	Sigma-Aldrich	Cat#N0640
IAA	Sigma-Aldrich	Cat#I2886
5-F-IAA	Goldbiotech	Cat#F-160-5
PEO-IAA	Sigma-Aldrich	Cat#CDS013406
Brassinolide	Sigma-Aldrich	Cat#B1439
Benzoid Acid (BA)	Sigma-Aldrich	Cat#242381
Experimental Models: Cell Lines		
<i>Arabidopsis</i> suspension culture (PSB-L)	N/A	N/A
Experimental Models: Organisms/Strains		
<i>Arabidopsis</i> : MAKR2-Ox1 (2x35Sprom::MAKR2-2xmCHERRY)	This manuscript	NASC #N2110134
<i>Arabidopsis</i> : MAKR2-Ox2 (2x35Sprom::MAKR2-mCITRINE)	¹⁷	NASC #N2110135
<i>Arabidopsis</i> : amiMAKR2	This manuscript	NASC #N2110142 and #N2110143
<i>Arabidopsis</i> : DR5::GUS	³⁹	N/A
<i>Arabidopsis</i> : DR5rev::GFP	⁴⁰	NASC #N9361
<i>Arabidopsis</i> : DR5::GUS;MAKR2-Ox1	This manuscript	N/A
<i>Arabidopsis</i> : DR5rev::GFP;MAKR2-Ox1	This manuscript	N/A
<i>Arabidopsis</i> : makr2-1 (crispr)	This manuscript	NASC #N2110144
<i>Arabidopsis</i> : MAKR2prom::MAKR2-tdYFP	This manuscript	NASC #N2110140
<i>Arabidopsis</i> : MAKR2prom::MAKR2-GUS	This manuscript	N/A
<i>Arabidopsis</i> : MAKR2prom::VENUS ^{NLS}	This manuscript	NASC #N2110141
<i>Arabidopsis</i> : MAKR2prom::MAKR2-2xmCHERRY	This manuscript	N/A
<i>Arabidopsis</i> : PIN2prom::PIN2-GFP	⁴¹	N/A
<i>Arabidopsis</i> : PIN2prom::PIN2-GFP;MAKR2-Ox1	This manuscript	N/A
<i>Arabidopsis</i> : PIN2prom::PIN2-GFP;amiMAKR2	This manuscript	N/A
<i>Arabidopsis</i> : ROP6prom::mCITRINE-ROP6 in rop6-2	²⁵	NASC #N2109740

(Continued on next page)

Continued

REAGENT or RESOURCE	SOURCE	IDENTIFIER
<i>Arabidopsis</i> : ROP6-Ox	27	N/A
<i>Arabidopsis</i> : MAKR2-Ox1;ROP6-Ox	This manuscript	N/A
<i>Arabidopsis</i> : TMK1-Ox (UBQ10prom::TMK1-2xmCHERRY)	This manuscript	NASC #N2110145
<i>Arabidopsis</i> : MAKR2-Ox2;TMK1-Ox	This manuscript	N/A
<i>Arabidopsis</i> : tmk1-1;tmk4	33	SALK_01360 Wiscseq_DsLox377-380D21.1
<i>Arabidopsis</i> : TMK1prom::2xmCHERRY ^{NLS}	This manuscript	N/A
<i>Arabidopsis</i> : MAKR2-Ox2;TMK1 ^{K616R} -Ox	This manuscript	N/A
<i>Arabidopsis</i> : MAKR2-mCit ^{GEGE}	This manuscript	NASC #N2110136 and #N2110137
<i>Arabidopsis</i> : MAKR2 ^{11Q} -mCit	This manuscript	NASC #N2110138 and #N2110139
Oligonucleotides		
Primer: MAKR2prom-topo F GTCTCTTTCAGTCATACCTCTCTCTAA	This manuscript	N/A
Primer: MAKR2prom-topo R TGTTGGGAAAGCCAGAATCA	This manuscript	N/A
Primer: MAKR2-B1 GGGGACAAGTTTGTACAAA AAGCAGGCTTAACCATGGAAGCTT TCAGTCTCCTAAC	This manuscript	N/A
Primer: MAKR-B2 GGGGACCACTTTGT ACAAGAAAGCTGGGTATTTCTCGTAA GAGGAAGGTTCACT	This manuscript	N/A
Primer: tdYFP-B2R GGGGACAGCTTTCTTG TACAAAGTGGCTATGGTATCCAAAGGT GAAGAAGACA	This manuscript	N/A
Primer: tdYFP-B3 GGGGACAACCTTTGTATAAT AAAGTTGCTCACTTATACAGTTCTG CCATCCCC	This manuscript	N/A
Primer: TMK1-B1 F GGGGACAAGTTT GTACAAAAAAGCAGGCTTAA CCATGAAGAAAAGAA GAACCTTTCTTCT	This manuscript	N/A
Primer: TMK1-B2 R GGGGACCACTTTG TACAAGAAAGCTGGGT ATCGTCCATCTACT GAAGTGAATGACT	This manuscript	N/A
Primer: tdYFP-B2R GGGGACAGCTTTCTTGTA CAAAGTGGCTATGGTAT CCAAAGGTGAAGAAGACA	This manuscript	N/A
Primer: tdYFP-B3 GGGGACAACCTTTGTA TAATAAGTTGCTCACTTATACAGTTCTG TCCATCCCC	This manuscript	N/A
Primer: TMK1prom-Fw GTATAGAAAAG TTGCTGTGGAATTTTA ATCTTAATTAAGGAAGC	This manuscript	N/A
Primer: TMK1prom-Rev TTTTGT ACAACTTGCAGCTTGA AGAAACAGAGGATTGAAGAAGAAACAG	This manuscript	N/A
Primer: p5'-open-Fw CCTCTGTTTCTTCAAGCTGCAAGTTT GTACAAAAAAGTTGAACG	This manuscript	N/A

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Continued

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Primer: p5'-open-Rev TTAAGATTAATAATCCACAGCAACTTTT CTATACAAAGTTGG	This manuscript	N/A
Primer: TMK1 ^{KD} (K616R)-F ATTGCGGTTAGGAGAATGGAGAATGGA GTTATTGCTGG	This manuscript	N/A
Primer: TMK1 ^{KD} (K616R)-R CCATTCTCCATTCTCCTAACCGCAAT CTTCGTTCCATC	This manuscript	N/A
Primer: TMK1-FL-B1 GGGGACAAGTTTG TACAAAAAGCAGGCTCAAT GAAGAAAAGAAGAACCCTTTC	This manuscript	N/A
Primer: TMK1-FL-B2-R GGGGACCAC TTTGTAAGAAAGCTG GGTCTCGTCCATCTACTGAAGTGAAT	This manuscript	N/A
Primer: TMK1-ΔKD-B2-R GGGGACCAC TTTGTAAGAAAGCTGGGTCGTTGTT AGTCACAGAACGAAG	This manuscript	N/A
Primer: F1 gRNA3 ATATATGGTCTCGATTGCTTTCAGTCTC CTTAACACTACGTT	This manuscript	N/A
Primer: F0 gRNA3 TGCTTTCAGTCTCCTTAACACTACGTTTGA GAGCTAGAAATAGC	This manuscript	N/A
Primer: R0 gRNA4 AACGCGGAGTAACGGCAAAGAGACA ATCTCTTAGTCGACTCTAC	This manuscript	N/A
Primer: R2 gRNA4 ATTATTGGTCTCGAAACGCGGAGTAAC GGCAAAGAGACAA	This manuscript	N/A
Primer: MAKR2 qPCR F CAACAAGATAGTATTCAAAGTGCCA	This manuscript	N/A
Primer: MAKR2 qPCR R GAGGAAGGTTCACTCACCAG	This manuscript	N/A
Primer: EF1 alpha qPCR F TGAGCACGCTCTTCTTGCTTTCA	This manuscript	N/A
Primer: EF1 alpha qPCR R GGTGGTGGCATCCATCTTGTTACA	This manuscript	N/A
Primer: GAPDH F GAATCCGAAGGCACAACTCAA	This manuscript	N/A
Primer: GAPDH R AAACTGTGCTCAATGCAA	This manuscript	N/A
Recombinant DNA		
Empty gateway destination vector: pB7m34GW	42	N/A
Empty gateway destination vector: pH7m34GW	42	N/A
Empty gateway destination vector: pLOK180_FR7m34GW	Gift from Lothar Kalmbach, Cambridge	N/A
Empty gateway destination vector: pMDC32	43	N/A
Empty gateway destination vector: pTNT- HA-GW	44	N/A

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REAGENT or RESOURCE	SOURCE	IDENTIFIER
Empty gateway destination vector: pIX:HALO:ccdB-CmR	45	ABRC #CD3-1742
Expression plasmid: pET28-duet	This manuscript	N/A
Entry gateway vector: <i>2x35prom/pDONR4-P1R</i>	46	NASC #N2106316
Entry gateway vector: <i>UBQ10prom/pDONR4-P1R</i>	46	NASC #N2106315
Entry gateway vector: <i>MAKR2prom/pENTR5'</i>	This manuscript	N/A
Entry gateway vector: <i>TMK1prom/pDONR4-P1R</i>	This manuscript	N/A
Entry gateway vector: MAKR2 ^{gnoSTOP} /pDONR221	17	N/A
Entry gateway vector: MAKR2 ^{wSTOP} /pDONR221	This manuscript	N/A
Entry gateway vector: TMK1 ^{noSTOP} /pDONR221	This manuscript	N/A
Entry gateway vector: TMK1 ^{K616R} ^{noSTOP} /pDONR221	This manuscript	N/A
Entry gateway vector: MAKR2(CDS) noSTOP/pDONR221	This manuscript	N/A
Entry gateway vector: MAKR2 ^{11Q} ^{noSTOP} /pDONR221	This manuscript	N/A
Entry gateway vector: 2xmCHERRY- 4xMyc/pDONR2R-P3	14	NASC #N2106292
Entry gateway vector: mCITRINE/ pDONR2R-P3	14	NASC #N2106288
Entry gateway vector: tdYFP/ pDONR2R-P3	This manuscript	N/A
Entry gateway vector: mCITRINE ^{GE} /pDONR2R-P3	17	N/A
Entry gateway vector: TMK1 ^{cyt} /pDONR221	This manuscript	N/A
Entry gateway vector: HSL1 ^{cyt} /pDONR221	47	ABRC #N1g28440Ze_K
Entry gateway vector: mCITRINE ^{wSTOP} /pDONR221	46	N/A
Entry gateway vector: TMK1 ^{kinase} /pDONR221	This manuscript	N/A
Destination expression vector: TMK1 ^{cyt} /pTNT-HA-GW	This manuscript	N/A
Destination expression vector: HSL1 ^{cyt} /pTNT-HA-GW	This manuscript	N/A
Destination expression vector: mCITRINE/ pIX-HA-GW	This manuscript	N/A
Destination expression vector: MAKR2/pIX-HA-GW	This manuscript	N/A
Expression vector: MAKR2-TMK1 ^{cyt} /pET28-duet	This manuscript	N/A
Expression vector: MAKR2-TMK1 ^{cytK616R} /pET28-duet	This manuscript	N/A
Destination vector: <i>UBQ10prom::TMK1-3HA/pB7m34GW</i>	This manuscript	N/A
Destination vector: <i>UBQ10prom::TMK1^{kinase}-3HA/pB7m34GW</i>	This manuscript	N/A

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Continued

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Destination vector: <i>2x35Sprom::MAKR2-2xmCHERRY/pH7m34GW</i>	This manuscript	N/A
Destination vector: <i>2x35Sprom::MAKR2-mCitrine/pB7m34GW</i>	This manuscript	N/A
Destination vector: <i>amiMAKR2/pMDC32</i>	This manuscript	N/A
Destination vector: <i>amiMAKR2/pB7WG2</i>	This manuscript	N/A
Destination vector: <i>MAKR2prom::MAKR2-2xmCHERRY/pH7m34GW</i>	This manuscript	N/A
Destination vector: <i>MAKR2prom::MAKR2-tdYFP/pB7m34GW</i>	This manuscript	N/A
Destination vector: <i>UBQ10prom::TMK1-2xmCHERRY-/pH7m34GW</i>	This manuscript	N/A
Destination vector: <i>UBQ10prom::TMK1^{K616R}-2xmCHERRY-/pH7m34GW</i>	This manuscript	N/A
Destination vector: <i>2x35S::MAKR2-mCit^{GEGE}/pB7m34GW</i>	This manuscript	N/A
Destination plasmid: <i>2x35S::MAKR2^{11Q}-mCit/pLOK180_pFR7m34GW</i>	This manuscript	N/A
Destination vector: <i>MAKR2prom::MAKR2-GUS</i>	This manuscript	N/A
Destination vector: <i>MAKR2prom::VENUS^{NLS}</i>	This manuscript	N/A
Destination vector: <i>TMK1prom::2xmCHERRY^{NLS}</i>	This manuscript	N/A
Software and Algorithms		
ImageJ	NIH ⁴⁸	https://imagej.nih.gov/ij/
R	R project	https://www.r-project.org/
Excel	Microsoft	https://www.microsoft.com/
Other		
Plant/Fungi RNA Purification Kit	Sigma-Aldrich	Cat#E4813
Reverse transcriptase SuperScript IV VILO Master Mix	ThermoFisher	Cat#11756050
Gateway BP clonase	ThermoFisher	Cat#11789100
Gateway LRll plus clonase	ThermoFisher	Cat#12538120
TnT Coupled Wheat Germ Extract System	Promega / ThermoFisher	Cat#L5030
Halo-tag magnetic beads	Promega / ThermoFisher	Cat#7281

RESOURCE AVAILABILITY

Lead Contact

Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, Yvon Jaillais (yvon.jaillais@ens-lyon.fr).

Materials Availability

There is no restriction of the material generated in this study (plasmids and *Arabidopsis* transgenic seeds).

Data and Code Availability

This study did not generate any unique datasets or code.

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Plant material and Growth conditions

In all the experiments, wild-type Col-0 ecotype of *Arabidopsis thaliana* was used as a control and all transgenic lines were produced in Col-0 background. Plants were grown in continuous light on MS supplemented with vitamins (Duchefa) and 1% sucrose. The following transgenic lines have been described before: 35S::GFP-ROP6 (ROP6-Ox);²⁷ DR5prom::GUS;³⁹ DR5rev-prom::GFP;⁴⁰ PIN2::PIN2-GFP;⁴¹ ROP6prom::mCitrine-ROP6g/rop6-2;²⁵ tmk1-1 (SALK_016360) and tmk1tmk4 mutants (tmk4: Wiscseq_DsLox377-380D21.1).³³

Plant Transformation and Selection

Each construct was transformed into C58 GV3101 *Agrobacterium tumefaciens* strain and selected on YEB media (5g/L beef extract; 1g/L yeast extract; 5g/L peptone; 5g/L sucrose; 15 g/L bactoagar; pH 7.2) supplemented with antibiotics (Spectinomycin, Gentamycin). After two days of growth at 28°C, bacteria were collected using a single-use cell scraper, re-suspended in about 200 mL of transformation buffer (10mM MgCl₂; 5% sucrose; 0.25% silwet) and plants were transformed by dipping. Plants from the Columbia-0 (Col0) accession were used for transformation.

Primary transformants (T1) were selected *in vitro* on the appropriate antibiotic/herbicide (glufosinate for mCITRINE, hygromycin for mCHERRY-tagged proteins or using the FastRed screening method). Approximately 20 independent T1s were selected for each line. In the T2 generation at least 3 independent transgenic lines were selected using the following criteria when possible: i) good expression level in the root for detection by confocal microscopy, ii) uniform expression pattern, iii) single insertion line (1 sensitive to 3 resistant segregation ratio). Lines were rescreened in T3 using similar criteria as in T2 with the exception that we selected homozygous lines (100% resistant/fluorescent). At this step, we selected one to three transgenic line(s) that was(were) used for further analyses and crosses.

METHOD DETAILS

Cloning and characterization of transgenic lines

The cloning for the transgenic lines production was performed using the multi-site gateway system (thermofisher).

Cloning of promoters into gateway entry vectors

The MAKR2 promoter was amplified from Col-0 genomic DNA using MAKR2prom-topoF/MAKR2prom-topoF (Key Resources Table) and cloned into the pENTR 5'-TOPO TA vector by TOPO TA Cloning (thermofisher) to give MAKR2prom/pENTR5'. The TMK1 promoter was amplified from Col-0 genomic DNA using TMK1prom-Fw and TMK1prom-Rev and cloned into pDONRP4-P1R using Gibson cloning, with pDONRP4-P1R having been amplified using the p5'-open-Fw and p5'-open-rev primers, to give TMK1prom/pDONRP4-P1R. 2x35Sprom/pDONRP4P1R and UBQ10prom/pDONRP4P1R were described previously.^{14,46}

Cloning of genes into gateway entry vectors

The MAKR2 genomic fragment was amplified from Col-0 genomic DNA using MAKR2-B1/MAKR2-B2 primers and introduced into the pDONR221 by BP recombination to give MAKR2^{noSTOP}/pDONR221. TMK1 was amplified from Col-0 cDNA using TMK1-B1/TMK1-B2 primers and recombined into pDONR221 by BP cloning to give TMK1^{noSTOP}/pDONR221. MAKR2(CDS)^{noSTOP}/pDONR221 was previously described.¹⁷ TMK1^{KD(K616R)}^{noSTOP}/pDONR221 was obtained by site directed mutagenesis by amplifying TMK1^{noSTOP}/pDONR221 with the TMK1^{KD}-K616F/TMK1^{KD}-K616R primer pair.

The MAKR2^{11Q} (CDS) no STOP sequence was synthesized by IDT technologies in the pUCIDT-AMP vector and subsequently recombined into pDONR221 by BP reaction to obtain MAKR2^{11Q} (CDS) no STOP/pDONR221. In the MAKR2^{11Q} sequence, the 11 positively charged residues (lysine -K- or arginine -R-) of the cationic region were mutated to the neutral amino acid glutamine (Q) as follows:

> MAKR2

780 gag aaa cga ttc gtg atg atg caa aag tac tta aag aag gta aaa cca ctt tac atc aga

E K R F V M M Q K Y L K K V K P L Y I R

gtt tca cgt cgt tac ggc gag aaa tta cga cac 870

V S R R Y G E K L R H

> MAKR2^{11Q}

780 gag caa caa ttc gtg atg atg caa cag tac tta cag cag gta caa cca ctt tac atc caa

E Q Q F V M M Q Q Y L Q Q V Q P L Y I Q

gtt tca caa cag tac ggc gag caa tta caa cac 870

V S Q Q Y G E Q L Q H

Cloning of artificial microRNAs into gateway entry vectors

Artificial microRNAs were designed using web microRNA designer (<http://wmd3.weigelworld.org/cgi-bin/webapp.cgi>). The following sequence was synthesized by IDT technologies and subsequently recombined into pDONR/Zeo by BP recombination to give ami-MAKR2/ pDONR/Zeo:

> amiMAKR2

acaagttgtacaaaaagcaggtcacaacacacgctcggacgcattacacatgttcatacacttaactcgtgtttgaattgatgttttaggaatatatgt**agatatg**taaa
gtggttttacctac**acaggtcgtgatgatgattcaattagcttccgactcattcatccaataaccgagtcgcca**aaattcaactagactcg**ttaaatgaatgaatgatcg**
gtagacaaattggatcattgattctctttgataggtaaaaccactttacata**ctctctttgtattccaattttcttgattaatcttctgcacaaaaacatgctgatccactaagtga**
catatatgctgcctcgtatatagttctggtaaaataacattttgggttatctttatgaaggcatcgccatgacccagcttctgtacaagtggt

Attb1 and Attb2 for gateway cloning

amiMAKR2/ pDONR/Zeo was transferred by LR recombination into pMDC32⁴³ or pB7WG2.⁴²

Cloning of reporters into gateway entry vectors:

tdYFP is a tandem dimer of dlanYFP (from *Branchiostoma lanceolatum*).⁴⁹ The following sequence was codon optimized for *Ara-*
bidopsis and synthesized by IDT (<https://eu.idtdna.com/site>):

> tdYFP:

atggtatccaaggtgaagaagacaatatggcatcattacctgcaacccatgagcttcacatctttggt
cgttaatggagtagattttgataggttggaagagggacaggttaaccctaacgacggttatgaagaact
taacctaaatctaccaagggagatctccaattctctcctggattttggcccacagattggttatggt
tttcatcagtatttgcatttcccgcaggaatgtctccattccaagcagccatgaagacggctcgggct
accaggttcacagaacaatgcaattgaagatggtgcaagtctcacttcaactatcgttatacgtatga
aggaagtcataataaaggcgaatttcaggtaaagggaactggatttctgcccacgagaccagttatgaca
aattcattgactgcagcagattggtgtgtgacaaagatgttatccaacgacaagactatcatctcaa
cttttgattggactacactacgggtaattgaaaaaggtatcaaacgacggcaagaaccacttacacatt
cgccaagccaatggcagcaaacataactaaagaccagccaatgttcgttttagaagacagagctcaag
cattcaaagactgaacttaactcaagaatggcaaaaagcattcaccgatgtgatgggtcatggaactg
gatctacgggatctggctcaagtgaacagcaagttctgaagataataatggcatcgctgcctgcaac
ccatgaactcatatatttgggtcttcaatggagtcgatttgacatggttaggacgaggtacaggtaat
cctaacgatgggtacgaggaggtgaacctaaagagtactaaggagacccctcagttcagtccttgattt
tagtaccgcaaatcggtatggttccaccagttattaccatttccggacggaatgtcgccttcaggc
ggcgatgaaagacggctcgtgatcaggttcatagaacctgaatttgaggacggagcactctctgacg
tccaactatagatatacttatgaaggctcgcacattaaaggagagttcaggtgaagggaactggttcc
ctgctgacggcctgtcatgacaaatagccttactgctcggattggtgtgttaccaaaatgctctaccc
taatgacaagactatcatcagtttactggtgacttatactactggaacgggaagcgatcatcagtc
acggcaagaactacatacacttgcacaaacctatggcagccaacatctgaaaaaccaacaaatgtttg
tgttcaggaagactgaactaaacactcaaaaaccgaactgaatttcaaagagtggcagaaagctttcac
agatgttatgggatggacgaactgtataagtga

The synthetic tdYFP gene was subsequently amplified using the tdYFP-B2R/tdYFP-B3 primer pair and cloned into pDONRP2R-P3 using BP recombination. mCITRINE/pDONRP2RP3, mCITRINE^{GE}/pDONRP2RP3, 2xmCHERRY-4xMyc/pDONRP2RP3, GUS/pDONRP2R-P3, VENUS^{NLS}/pDONR221 and mock/pDONRP2RP3 were described previously.^{14,17,46,50}

All primers used for cloning are indicated in the [Key Resources Table](#).

Destination vectors and plant transformation

Arabidopsis stable transformation and selection were performed as described.⁵¹ Final destination vectors for plant transformation were obtained using the LR recombination system (<http://www.thermofisher.com/>) using the pB7m34GW⁴² (basta resistant), pH7m34GW⁴² (hygromycin resistant), pB7WG2 (basta resistant),⁴² pLOK180_pFR7m34GW (gift from Lothar Kalmbach, Cambridge, similar backbone as pB7m34GW but with the basta resistance replaced by a FASTRED cassette for selection of transgenic seeds via red fluorescence) or pMDC32⁴³ (hygromycin resistant) destination vectors.

The following Gateway LR reactions were set-up to generate the corresponding destination vectors (the name of the corresponding transgenic line is highlighted in bold):

2x35Sprom::MAKR2-2xmCHERRY/pH7m34GW (**MAKR2-Ox1**) was obtained by recombining 2x35Sprom/pDONR4-P1R, MAKR2gnoSTOP/pDONR221, 2xmCHERRY-4xMyc/pDONR2R-P3, and pH7m34GW.

2x35Sprom::MAKR2-mCITRINE/pB7m34GW (**MAKR2-Ox2**) was obtained by recombining 2x35Sprom/pDONR4-P1R, MAKR2gnoSTOP/pDONR221, mCITRINE/pDONR2R-P3, and pB7m34GW.

2x35Sprom::amiMAKR2/pMDC32 (**amiMAKR2.1** and **amiMAKR2.3**) 2x35Sprom::amiMAKR2/pB7WG2 (**amiMAKR2.2**) were obtained by recombining amiMAKR2/pDONR221, with pMDC32 or pB7WG2, respectively.

MAKR2prom::MAKR2-2xmCHERRY/pH7m34GW (**MAKR2::MAKR2-2Ch**) was obtained by recombining MAKR2prom/pDONR4-P1R, MAKR2gnoSTOP/pDONR221, 2xmCHERRY-4xMyc/pDONR2R-P3, and pH7m34GW.

MAKR2prom::MAKR2-tdYFP/pB7m34GW (**MAKR2::MAKR2-tdYFP**) was obtained by recombining MAKR2prom/pDONR4-P1R, MAKR2gnoSTOP/pDONR221, tdYFP/pDONR2R-P3, and pB7m34GW.

MAKR2prom::MAKR2-GUS/pK7m34GW (**MAKR2::MAKR2-GUS**) was obtained by recombining MAKR2prom/pDONR4-P1R, MAKR2gnoSTOP/pDONR221, GUS/pDONR2R-P3, and pK7m34GW.

MAKR2prom::VENUS^{NLS}/pB7m34GW (**MAKR2::VENUS^{NLS}**) was obtained by recombining MAKR2prom/pDONR4-P1R, VENUS^{NLS}/pDONR221, mock/pDONR2R-P3, and pB7m34GW.

TMK1prom::CHERRY^{NLS}/pH7m34GW (**TMK1::2Ch^{NLS}**) was obtained by recombining TMK1prom/pDONR4-P1R, mCHERRYnoSTOP/pDONR221, mCHERRY^{NLS}/pDONR2R-P3, and pH7m34GW.

UBQ10prom::TMK1-2xmCHERRY/pH7m34GW (**TMK1-Ox**) was obtained by recombining UBQ10prom/pDONR4-P1R, TMK1noSTOP/pDONR221, 2xmCHERRY-4xMyc/pDONR2R-P3, and pH7m34GW.

UBQ10prom::TMK1^{K616R}-2xmCHERRY/pH7m34GW (**TMK1^{K616R}-Ox**) was obtained by recombining UBQ10prom/pDONR4-P1R, TMK1^{K616R}noSTOP/pDONR221, 2xmCHERRY-4xMyc/pDONR2R-P3, and pH7m34GW.

2x35Sprom::MAKR2-mCITRINE^{GEGE}/pB7m34GW (**MAKR2-mCit^{GEGE}**) was obtained by recombining 2x35Sprom/pDONR4-P1R, MAKR2(CDC)/pDONR221, mCITRINE^{GEGE}/pDONR2R-P3, and pB7m34GW.

2x35Sprom::MAKR2-mCITRINE^{GEGE}/pLOK180_pFR7m34GW (**MAKR2^{11Q}-mCit**) was obtained by recombining 2x35Sprom/pDONR4-P1R, MAKR2^{11Q}(CDC)/pDONR221, mCITRINE/pDONR2R-P3, and pLOK180_pFR7m34GW.

Cloning and characterization of *makr2* crisp allele

The *makr2-1* crisp allele (Col0) was generated using the egg cell-specific promoter (*pEC1.2*) CRISPR/Cas9 system described in Wang et al.⁵² Two single guide RNAs (sgRNA) were originally used in the construct: *sgRNA3* 5'-CTTTCAGTCTCCTTAACCTAC-3' and *sgRNA4* 5'-TCTCTTTGCCGTTACTCCG-3' but mutations were only found in the *sgRNA3* target sequence. Primers containing the sgRNA sequences and the BsaI restriction site were designed (F1 *gRNA3*: ATATATGGTCTCGATTGCTTTCAGTCTCCTTAAC TACGTT; F0 *gRNA3*: TGCTTTCAGTCTCCTTAACCTACGTTTTAGAGCTAGAAATAGC; R0 *gRNA4*: AACGCGGAGTAACGGCAAAGA GACAATCTCTTAGTCTGACTCTAC; R2 *gRNA4*: ATTATTGGTCTCGAAACGCGGAGTAACGGCAAAGAGACAA) and a fourth primer PCR was performed using the *pCBC-DT1T2* vector as a template. The PCR fragment containing the sgRNAs was introduced to the binary vector *pHEE401E* by golden gate cloning using BsaI restriction sites. Plants were transformed and T1 plants selected by hygromycin resistance. Plants carrying mutations were selected by sequencing and, homozygous mutant plants depleted of the T-DNA were counterselected in T2. The *makr2-1* mutant carries a frameshift mutation (1 nucleotide (A) insertion) at position 25 relative to the *MAKR2* start ATG. The resulting predicted short *MAKR2* protein comprises 37 amino acids. The first 8 amino acids correspond to *MAKR2* wt sequence, and amino acids 9 to 37 represent an aberrant amino acid sequence.

Gravitropism experiments

The horizontal gravitropic index was quantified using ImageJ as indicated in Platre et al.²⁵ and Grabov et al.⁵³ Plants were grown on plates with a 45° angle. The horizontal growth index was calculated on 7-day-old seedlings using the “segment line” tool on FIJI. Briefly, to calculate the gravitropic indexes, two lengths are considered, L and Lx.⁵³ L is the total length of the roots (from base of hypocotyl to root tip), while Lx is the abscissa of the root tip (considered from the point of view of the base of hypocotyl). The horizontal growth index (HGI) corresponds to the ratio Lx/L. The experiments were performed 3 times and at least 21 plants of each genotype were quantified in every independent experiment.

Gravitropism experiments were performed either on 5 to 6-days-old seedlings grown in MS media containing 1% sucrose. In order to align the roots, one hour prior to the experiment, seedlings were transferred in a new vertical plate. An angle of 135° was applied in darkness and the same plates were scanned after 3 or 4 hours, 6 hours, 12 hours, 24 hours or 48 hours. The angle of the root bending at each time point was quantified using ImageJ.⁴⁸ The experiment was performed 3 independent times and at least 30 plants of each genotype were quantified.

Kinetics of the PIN2 dynamics during gravitropism was performed as follows: 5-days-old seedlings were transferred and aligned in new 1/2 MS Petri dishes one hour prior to the experiment. In this case, an angle of 90° was applied and fluorescent images were taken every five minutes during six hours. The angle of the root bending was quantified using ImageJ.

Quantitative RT-PCR (qPCR)

Total RNA was isolated using the Plant/Fungi RNA Purification Kit (sigma) and quantified using Nanodrop 1000 (Thermo Scientific, <http://www.nanodrop.com/>). 1 µg of total RNA was reverse transcribed and amplified using the SuperScript IV VILO Master Mix (ThermoFisher). Transcript levels were measured by qPCR using amplified cDNA. The relative amount of each transcript was calculated with the 2^{-ΔCT} method⁵⁴ using *EF1-alpha* and *GADPH* transcripts as housekeeping for data normalization. Each experiment was performed in at least three biological replicates. The qPCR primers used are described in the table below.

Drugs and hormones treatments

Drugs and hormones treatments were performed in liquid 1/2 MS on 5 days-old-seedlings. Cycloheximide (CHX) (stock 50 mM in DMSO) and MG-132 (stock 25 mM in DMSO) treatments were carried out at 50 µM during two hours before the NAA treatment.

NAA treatments were done at 1 µM or 10 µM during 5 or 30 minutes, as indicated in the experiment. The auxin analogs 5-F-IAA (stock 10 mM in ethanol), PEO-IAA (stock 10 mM in ethanol) and benzoic acid (BA) (stock 10 mM in ethanol) were diluted in 1/2 MS at 10 µM during 30 minutes. Brassinolide (stock 10 mM in DMSO) treatments were performed at 10 µM during 30 minutes.

Each treatment was repeated in at least three independent experiments and quantifications were performed on at least 14 roots. The number of cells quantified is indicated in the corresponding graphs.

Microscopy

The experiments were performed using either the LSM710 confocal microscope using a 40X Plan-apochromatic objective (numerical aperture 1.2, oil immersion) (Zeiss) or inverted Zeiss microscope (AxioObserver Z1, <http://www.zeiss.com/>) equipped with a spinning disk module (CSU-W1-T3, Yokogawa, <http://www.yokogawa.com/>) and a ProEM+ 1024B camera (PrincetonInstrument, <http://www.princetoninstruments.com/>) using a 40X C-Apochromat objective (numerical aperture 1.1, water immersion).

tdYFP and mCitrine were excited at 512 nm and mCherry was excited at 550 nm. FM4-64 staining were performed as described in Marquès-Bueno et al.⁴⁶

MAKR2-Ox1 imaging was performed using a FV 1000 confocal microscope (Olympus, Tokyo, Japan) mCherry was excited at 550 nm and using 40X oil objective.

Microfluidic experiments were performed using a vertical ZEISS LSM700 confocal microscope with a 20x/0.8 Plan-Apochromat M27 objective.⁵⁵ In this case, tdYFP was excited at 488 nm and images were taken every minute, during six minutes. First, three images of roots in liquid MS media without hormones were acquired and afterward NAA 10 μM or IAA 10 μM or benzoic acid 10 μM were added and three additional pictures were taken.

The kinetics of the PIN2 dynamics was performed using a vertically mounted Olympus MVX10 macroview fluorescence stereomicroscope,²⁸ setting up the automated filterwheels to allow fluorescence imaging for GFP and using 5x magnification. (<http://www.olympusamerica.com>). For the time lapse imaging, "Process Manage" function in Cellsens Dimension software (Olympus) was used. After finishing the acquisition, the images were saved as a video and analyzed using ImageJ software.

6-days-old seedlings of *MAKR2prom::MAKR2-GUS*, *DR5prom::GUS* or *MAKR2ox1-DR5prom::GUS* were fixed in cold 90% acetone on ice during 20 minutes, then washed three times with 50 mM NaHPO₄ buffer (pH 7.2) and stained over night with 2 mM X-Gluc staining buffer at 37 °C in the darkness. Afterward, roots were washed with ethanol series from 30%, 50%, 70% 20 min each step and then finally 100%. Finally, the roots were mounted with chloralhydrate and pictures were obtained using the Leica DM6 (Leica, Germany) epifluorescence vertical microcroscope equipped with a Normansky optics and using either 20x or 40x magnification. Quantifications were done using region-of-interests drawn in the upper and lower part of the root tip and GUS/GFP intensity were quantified using the mean intensity tool from ImageJ.

In the case of *DR5prom::GUS*, the analyses were made on 60 roots and three independent experiments (20 roots per replicate), and 42 roots and three independent experiments (14 roots per replicate) for *DR5prom::GFP*.

In vitro Halo-pull down

The cytoplasmic domains of TMK1 and HSL1 cloned into pDONR221 was recombined by LR reaction into pTNT-HA-GW.⁴⁴ The HSL1^{cyt} clone was ordered to ABRC.⁴⁷ mCITRINEwSTOP/pDONR221 and MAKR2wSTOP/pDONR221 were recombined by LR reaction into pIX:Halo:ccdb.⁴⁵ The TnT Coupled Wheat Germ Extract System from Promega were used for cell free expression. 4 μg of both plasmids were added to the reaction mix following the commercial protocol. The reactions were incubated at 25°C for 2 hours. The equivalent of 10 μl of dry Halo-tag magnetic beads from Promega were added and the samples were incubated at room temperature for 30 minutes on an orbital shaker. The supernatant was discarded and the beads were incubated for 3 minutes in PBS + 0.05% Tween20. The solution was discarded. This wash was repeated 3 times. The beads were then resuspended in SDS-PAGE loading buffer, separated by SDS-PAGE and subjected to immunoblotting using anti-Flag antibody (clone M2, F1804, Sigma) and Anti-HA antibody (12CA5, Sigma-Aldrich).

Co-expression, purification and mass-spectrometry

To generate the backbone plasmid for co-expression in bacteria, *pET28a+* and *pACYC-duet* were digested using Xba1 and Xho1. The MCS cassette of *pACYC-duet* was inserted into *pET28a+* leading to the generation of *pET28-duet*. *MAKR2* and *TMK1* were amplified by PCR from Col-0 cDNA. The tags and restriction sites were added by PCR. *MAKR2* PCR fragment was cloned into *pET28-duet* between Nde1 and Xho1 and *TMK1* PCR fragment between Nhe1 and Not1. Mutation to generate a dead kinase version of TMK1 was done by PCR. The resulting plasmids were transformed in *Escherichia coli* BL21 (DE3) Rosetta 2 from Novagen that were spread on LB + 100 μg/μL kanamycin plates. Several colonies were picked and transferred into 100ml of 2xTY + 100 μg/μL kanamycin and incubated overnight at 37°C under 180rpm shaking. The day after, a 250ml culture of 2xTY + 100 μg/μL kanamycin was inoculated with the preculture to reach a 0.1 OD₆₀₀ and incubated at 37°C under 180 rpm shaking until OD₆₀₀ 0.6. Then 0.5 mM IPTG was added and the samples were incubated for 75 minutes at 37°C. The cultures were pelleted through a 5000 rcf centrifuge for 20 minutes. The pellets were resuspended in 15 mL of resuspension buffer (20 mM Tris pH8, NaCl 150mM and 20mM imidazole). The resuspended pellets were flash frozen and stored at -20°C. The pellets were unfrozen, 5 mM beta-mercaptoethanol was added, then sonicated and centrifuged at 15000 rcf for 40 minutes at 4°C. The soluble fraction was applied on 0.5 mL of IMAC-nickel resin. The resin was washed with 10 column volumes of resuspension buffer, then 5 column volumes of resuspension buffer + 40 mM imidazole. The proteins were eluted within 2 column volumes of Tris 20mM pH8, NaCl 150mM and 400 mM imidazole. The samples were analyzed by mass spectrometry (ESI/MS/MS) by PSF platform of SFR Biosciences (UMS3444/CNRS, US8/Inserm, ENS de Lyon, UCBL).

Co-immunoprecipitation Assay

Co-immunoprecipitation assays were performed in the *Arabidopsis* suspension culture (PSB-L) protoplast system. Protoplast were isolated and transfected or co-transfected with 15 μ g of plasmid DNA (*UBQ10::TMK1-3HA*, *UBQ10::TMK1^{Δkinase}-3HA*, *2x35S::MAKR2-mCitrine*) according to the method described previously.⁵⁶ After transfection, protoplasts were incubated for 16 hours in the dark to allow gene expression, treated with either DMSO or 1 μ M IAA dissolved in DMSO for 1 hour, collected and frozen in liquid nitrogen. For coimmunoprecipitation (Co-IP) assay, 5×10^5 transfected protoplasts were lysed with 50 μ l of extraction buffer (50 mM Tris HCl pH 7.5, 150 mM NaCl, 10 mM MgCl₂, 1% (w/v) Triton X-100, 5 mM DTT, PhosSTOP phosphatase inhibitor cocktail (Roche), 1x EDTA free-Complete Protease Inhibitor Cocktail (Roche)). After vortexing vigorously for 30 s, the samples were centrifuged at $14,000 \times g$ for 10 min at 4°C and supernatants were collected. Prior to the Co-IP assay, 25 μ l of collected supernatants were analyzed by immunoblot assay to determine Co-IP input. To perform the Co-IP assay, 75 μ l of extraction buffer without Triton X-100 were added to remaining 25 μ l of supernatants, followed by incubation with GFP-Trap Magnetic Agarose (ChromoTek) for 1 hour at 4°C, three washing steps with washing buffer (50 mM Tris HCl pH 7.5, 200 mM NaCl, 1 mM EDTA, 0.1% Triton X-100), once with 50 mM Tris HCl pH 7.5 and elution step with 2x-SDS-sample buffer for 10 min at 95°C. Samples were separated by SDS-PAGE and subjected to immunoblotting using anti-GFP-HRP antibody (Miltenyi Biotec) and Anti-HA-Peroxidase, High Affinity (3F10) (Roche).

To generate *UBQ10::TMK1-3HA*, *UBQ10::TMK1^{Δkinase}-3HA* constructs, the *TMK1* full length/ *TMK1^{Δkinase}* (amino acid 1-587) cDNA without stop codon were amplified by RT-PCR from WT total RNA using *TMK1-FL-B1-F* and *TMK1-FL-B2-R/TMK1-ΔKD-B2-R* primers, [Key Resources Table](#)), respectively, and inserted into the pDONR221 vector by BP recombination reaction. Next, *pDONR P4-P1R-UBQ10prom*, *pDONR P2R-P3-3xHA* and *pDONR221-gTMK1/gTMK1^{Δkinase}*, were cloned into *pB7m34GW* vector, respectively, by MultiSite Gateway LR recombination reaction.

Image Quantification

The quantification of PIN2-GFP fluorescence during gravitropism was performed as followed. Using ImageJ software (<https://imagej.nih.gov/ij/>), we draw a line (1px in width and 200px in length) on both sides of the root (upper and lower side) at each time point and used the mean intensity fluorescence tool from ImageJ for quantification. Afterward, the ratio between t/t₀ (t₀ being the initial fluorescence intensity at the beginning of the experiment) was applied to be able to follow the difference of fluorescence. Each graph in [Figure 2](#) were made from three independent roots.

Quantifications of MAKR2 translocation from the PM to the cytoplasm was carried as described in Simon et al.¹⁷ Briefly, we scored, in a double-blind set-up, the number of cells in which MAKR2-tdYFP or MAKR2-mCit was associated or not with the plasma membrane. A cell was counted as positive for plasma membrane labeling when the cell outline was at least twice as fluorescent as the cytosol. All the results are expressed as a percentage of the number of cells with MAKR2 at the plasma membrane over the total number of cells counted. The number of cells (n) counted in each case is indicated on the corresponding figure. Each experiment was repeated three time and at least 14 independent roots were counted.

For the microfluidic experiments, the quantification was performed by drawing a line at the plasma membrane and in the cytoplasm and quantifying the mean average intensity of both (ImageJ). With these values, the ratio of the fluorescence between the plasma membrane and the cytoplasm was applied and represented in the graphs.

QUANTIFICATION AND STATISTICAL ANALYSIS

Phenotyping

For the horizontal gravitropic index and the kinetics of the gravitropic response, a linear model was fitted on measurements from wild-type plants and the different mutants using *lm()* function from stats package available in R software (<https://www.r-project.org/>). This model estimates a weight for each variable (wild-type and mutant plants) and the associated probability that such weight is different from zero based on a t test. The probability derived from the t test is the p value in this comparison and significant differences were considered when the p value was lower than 0.05.

Cell biology experiments

For translocation of MAKR2 from the plasma membrane to the cytoplasm a pairwise comparison between mock plants and plants subjected to different treatments was performed using a t test with Welch correction to account for unequal variances using R software (<https://www.r-project.org/>). The probability derived from the t test is the p value in this comparison and significant differences were considered when the p value was lower than 0.01.

All the graphs were drawn using excel software (Microsoft, <https://products.office.com/>) except for the boxplots which were drawn either with excel or the R software (<https://www.r-project.org/>).